

# Metabarcoding Aquatic Microbial Communities at Clove Lakes Park, Staten Island Veronica Frid, Vivian Lam, Jenny Li & Tyler Clabby

### Abstract

We aim to measure the fluctuations in bacteria populations over time, specifically, as the weather grows warmer. We collected water samples from a small stream in Clove Lakes Park on Staten Island, NY. Utilizing centrifugation methods to pellet microorganisms from the water samples, we were able to extract DNA and sequence it to assess the estimated abundance of the microbial populations residing in the samples. By viewing the effects of increasing temperatures on the amounts of bacteria concentrated in water, we can draw conclusions as to how environmental factors influence the composition of aquatic microbial communities.

## Introduction

- Clove Lake is a freshwater lake located on Staten Island
- We sought to assess the microbial makeup of Clove Lake through metabarcoding over two months in Spring
- We hypothesized that the warmer temperatures at the second collection date will impact microbial diversity (Currie, et al, 2017)
- Previous research (Tsuji et al., 2017) stated that bacterial abundance is higher at higher water temperatures
- Ambient air temperatures leading up to sample collection, as well as precipitation were recorded to see if there was a correlation between microbial diversity and weather
- We also hypothesized that human factors can also have an impact on microbial makeup. For example, salt is used during winter months to prevent ice. Runoff from paved areas to Clove Lake could impact bacterial makeup of the water

# Materials & Methods

- Water samples from Clove Lake were collected on March 9th and April 6th, 2024.
- The samples were centrifuged multiple times to concentrate bacterial cells, and DNA extraction was performed using a Qiagen PowerSoil kit.
- Metabarcoding with 16s primers was conducted, followed by PCR and gel electrophoresis to check for DNA
- Illumina Amplicon sequencing was performed by GeneWiz
- Data analysis was done by using the ScienSano (Belgian Public Health Department) 16s pipeline on Galaxy
- The results included NCBI blast summaries and a Krona pie chart
- Additionally, weather data from Weather Underground was used for analysis on how environmental factors such as precipitation may impact the bacterial populations in the aquatic environment of Clove Lake.

### Acknowledgements

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(Staten Island Technical HS, Brooklyn Technical HS, New Dorp HS)

Table 1- Weather events for March 6-9 2024 (Source- Weather

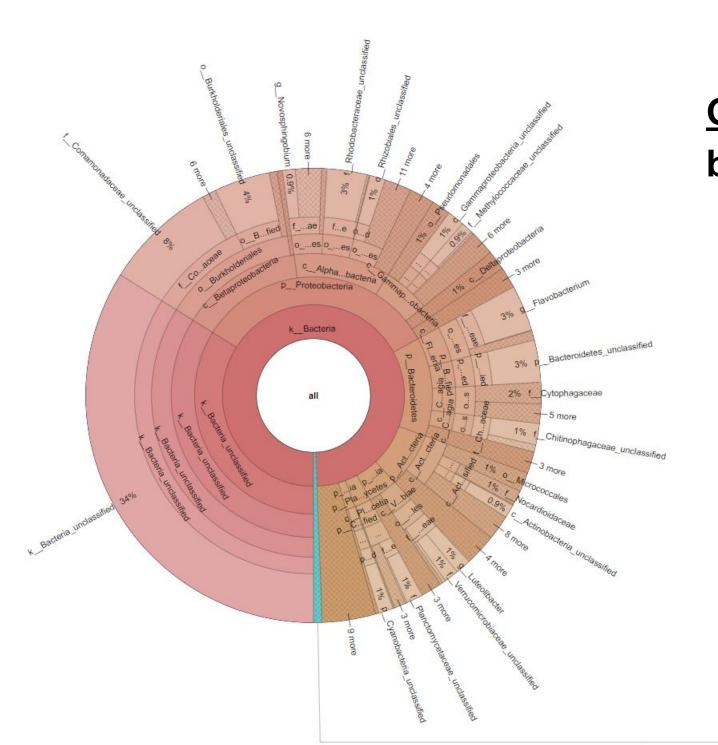
Underground Station KNYNEWY01176)				
Date	High temperature		Precipitation	
March 6th	48.2°F	45.2°F	1.56 inches	
March 7th	57.6°F	51.5° F	0.11 inches	
March 8th	57°F	38.7°F	0.0 inches	
March 9th* Sample collection	46.6°F	39.7° F.	1.30 inches	
date				
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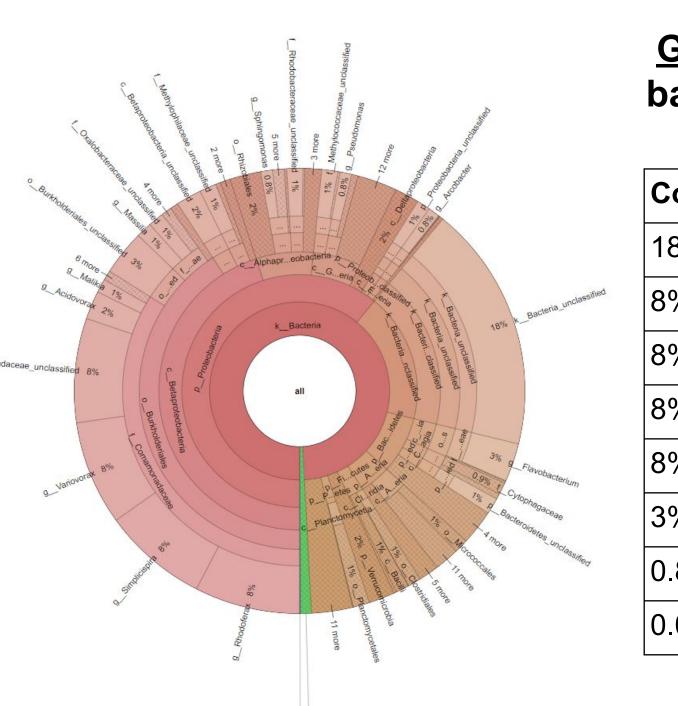
 

 Table 2- Weather events for April 3-6 2024 (Source- Weather

**Underground Station KNYNEWY01176)** 

Date	High temperature	Low temperature	Precipitation
April 3rd	43.5°F	40.3°F	1.42 inches
April 4th	51.4°F	35.6°F	0.33 inches
April 5th	52.7°F	36.7°F	0.00 inches
April 6th* Sample collection date	55.6°F	37.8°F	0.00 inches





### **<u>Graph 1</u>**: Krona table of largest Genera bacteria compositions + Archaea

Composition	Genera
34%	Unclassified bacteria
8%	Comamonadaceae
4%	Burkholderiales
3%	Rhodobacteraceae
3%	Flavobacterium
3%	Bacteroidetes
2%	Cytophagaceae
0.6%	Archaea

Archaea 0.6%

### **<u>Graph 2</u>**: Krona table of largest Genera bacteria compositions + Archaea

omposition	Genera
3%	Unclassified bacteria
%	Comamonadaceae
%	Variovorax
%	Simplicispira
%	Rhodoferax
%	Burkholderiales
8%	Archaea
05%	Unknown
05%	Unknown

### **Results and Discussion**

- 7°F increase
- eliminated.
- der Loos et al., 2020).

# References

2017, www.frontiersin.org/articles/10.3389/fmicb.2017.01599/full environmental DNA and its relation to bacterial abundance. PLoS ONE 12(4): e0176608. https://doi.org/10.1371/journal.pone.0176608 methodology involved." Molecular Ecology, vol. 30, no. 13, 29 Aug. 2020, pp. 3270–3288, https://doi.org/10.1111/mec.15592. Feb;52(2):73-116. doi: 10.1139/w05-147. PMID: 16541146. Temperature and microbial growth | Microbiology. (n.d.). /#:~:text=Psychrophiles%20grow%20best%20in%20the,to%20about%2045%20%C2%B0C https://dnr.wisconsin.gov/topic/Stormwater/learn\_more/problems.html



### The Pinkerton Foundation

• Over time, unclassified bacteria percentages decreased while the proportion of Camamonadaceae increased. We also saw changes in the percentages of Archaea DNA

 genera Variovorax, Simplicispira, and Rhodoferax all individually increased to 8% (a sum of 24%) by 4/6/2024 from a total 8% of unclassified bacteria under the Comamondacaeae family on 3/9/2024. This points to the conclusion that these genera prefer warmer temperatures over colder ones.

• Archaea are a separate kingdom from bacteria, but are also single-celled prokaryotes. They thrive in warm environments (Chaban B et al., 2006). This supports their increase from 0.6% in March to 0.8% in April, since the weather was warmer in April • Averaging temperatures from 3/9/24 and 4/6/24, we can see a

• Cytophagaceae and Burkholderiales genera decreased, from 2% and 4%, to 0.9% and 3%, respectively. Although Burkholderiales decreased by only one percent, which could be considered negligible, Cytophagaceae was nearly completely

• This could indicate that these bacteria prefer colder climates • These results show bias towards bacteria over Archaea species. There is also a bias towards the reference genomes available on NCBI. Other strains of common species could have been present but unable to be speciated on NCBI (van

Romero PE, Calla-Quispe E, Castillo-Vilcahuaman C, Yokoo M, Fuentes-Rivera HL, Ramirez JL, et al. (2021) From the Andes to the desert: 16S rRNA metabarcoding characterization of aquatic bacterial communities in the Rimac river, the main source of water for Lima, Peru. PLoS ONE 16(4): e0250401. https://doi.org/10.1371/journal.pone.0250401 Currie, Ashleigh R., et al. "Marine Microbial Gene Abundance and Community Composition in Response to Ocean Acidification and Elevated Temperature in Two Contrasting Coastal Marine Sediments." Frontiers, Frontiers, 7 Aug

Clove Lakes Park : NYC Parks, www.nycgovparks.org/parks/clove-lakes-park. Accessed 4 Jan. 2024. Tuhus, Melinda. "The Science of Litter." Urban Resources Initiative, Yale, 20 June 2020, uri.yale.edu/science-litter. Shahraki, Abdolrazagh Hashemi, et al. "Assessing High-Throughput Environmental DNA Extraction Methods for Meta-Barcode Characterization of Aquatic Microbial Communities." Journal of Water and Health, IWA Publishing, 1 Feb. 2019, iwaponline.com/jwh/article/17/1/37/64513/Assessing-high-throughput-environmental-DNA. Tsuji S, Ushio M, Sakurai S, Minamoto T, Yamanaka H (2017) Water temperature-dependent degradation of

Van der Loos, Luna M., and Reindert Nijland. "Biases in bulk: DNA metabarcoding of marine communities and the

Chaban B, Ng SY, Jarrell KF. Archaeal habitats--from the extreme to the ordinary. Can J Microbiol. 2006

https://courses.lumenlearning.com/suny-microbiology/chapter/temperature-and-microbial-growth

Why is Storm Water Runoff a Problem? | LEARN MORE ABOUT STORM WATER RUNOFF | Wisconsin DNR. (n.d.).