

# DNA Barcodes of Algae in Nutrient-Rich vs Nutrient-Poor NYC Waters

Mastura Akter<sup>1</sup>, Kripa Das<sup>2</sup>, Victoria Dubien<sup>2</sup>, Dr. Ellen Jorgensen<sup>3</sup>

Hillcrest High School<sup>1</sup>, Queens High School for the Sciences<sup>2</sup>, Biotech Without Borders<sup>3</sup>

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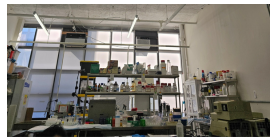
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## Abstract

New York City's coastal waters vary from nutrient-rich urban environments to more ocean-flushed conditions, providing an opportunity to study differences in algal diversity. This study compared algal communities from **Rockaway Beach, Sunset Cove, and the Hudson River** using **DNA metabarcoding of the chloroplast rbcL gene**. Six samples were collected, including both **surface water and rock biofilm brush-scrape samples**. Results showed that **brush-scraped biofilms contained significantly greater diversity than bulk-water samples**, recovering a wider range of diatoms and microalgae. In contrast, water samples were largely dominated by a single macroalgal taxon such as **Ulva or Fucus**. Overall, the findings suggest that **sampling method had a stronger effect on detected diversity than nutrient status**, emphasizing the importance of including substrate biofilms in environmental DNA biodiversity surveys.

## Introduction

- NYC coastal waters range from:
  - Nutrient-rich urban estuaries
  - To ocean-influenced oligotrophic environments
- Example:
  - Jamaica Bay receives high nitrogen input
  - Rockaway Beach experiences strong tidal flushing
- Traditional algal surveys:
  - Chlorophyll measurements
  - Microscopy
  - Often fail to identify species accurately
- DNA metabarcoding enables identification of multiple taxa from environmental DNA
- Study used the chloroplast rbcL barcode gene
- Research Question:
  - Do nutrient-rich and nutrient-poor waters show distinct algal diversity patterns?
- Initial Hypothesis:
  - Nutrient-rich waters would show:
    - Lower diversity
    - Pollution-tolerant algae dominance
  - Nutrient-poor waters would show:
    - Higher species richness



## Materials & Methods

### Sample Collection

- Sites sampled:
  - Rockaway Beach
  - Sunset Cove
  - Hudson River
- Two sample types collected:
  - Bulk-water samples
  - Brush-scrape biofilm samples
- Biofilms scraped from rocks using a sterile toothbrush
- Water filtered through a 0.45 µm filter

### DNA Extraction

- DNA extracted using:
  - Zymo Quick-DNA Fungal/Bacterial Microprep Kit
- DNA concentration measured using:
  - Qubit fluorometry

### PCR Amplification

- Target gene:
  - chloroplast rbcL
- Used:
  - Diatom-specific rbcL primers
- PCR products verified with:
  - 1% agarose gel electrophoresis

### Sequencing & Analysis

- Sequenced using:
  - Illumina paired-end 250 bp
- Reads:
  - Quality filtered
  - Clustered into OTUs
- Taxonomic identification performed using:
  - BLASTn against GenBank

## Results

We obtained 5 usable samples (A1, A2, A4, A5, A6); A3 (Hudson brush) produced no amplifiable DNA. After filtering, A1, A2, A4, and A5 each had ~5,000 reads, A6 had ~2,727, and A3 had ~1,017.

A1 (Rockaway-water): dominated by *Ulva lactuca*; 9 clusters.

A2 (Hudson-water): dominated by *Fucus spiralis*; 14 clusters.

A3 (Hudson-brush): no PCR product.

A4 (Rockaway-brush): most diverse sample (29 clusters), including *Ulva smaragdina* and many diatoms (*Navicula*, *Licmophora*, *Amphora*).

A5 (Sunset-water): dominated by *Strombidium* and *Monostroma*; 11 clusters.

A6 (Sunset-brush): contained diatoms (*Navicula*, *Amphora*) and other microalgae; 11 clusters.

Brush samples showed greater OTU diversity than water samples (e.g., A4: 29 clusters vs. A1: 9), while water samples were dominated by a single macroalga (*Ulva* or *Fucus*).

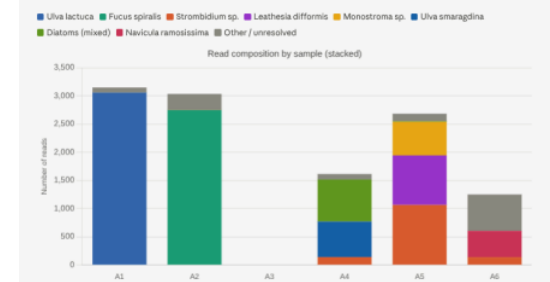
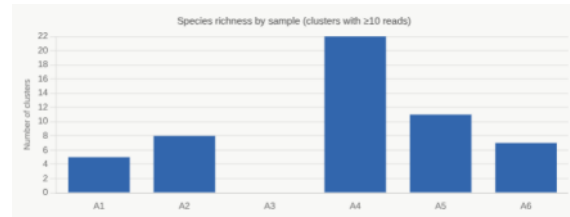


Table 1 and 2 (left to right) Summary of sequencing results by sample. Total reads = reads after quality filtering. Clusters (≥10 reads) = number of OTUs meeting the ≥10-read threshold. "Dominant taxa" are those with the highest read counts. Samples A1–A6 correspond to: A1=Rockaway-water, A2=Hudson-water, A3=Hudson-brush, A4=Rockaway-brush, A5=Sunset-water, A6=Sunset-brush.

## Discussion

- Water samples were mainly dominated by:
  - Ulva*
  - Fucus*
- Brush-scraped samples contained:
  - More diatoms
  - Green algae
  - Other microalgae
- Sampling method affected diversity results more than nutrient level
- Rock biofilms contained taxa not found in surface water
- Findings support previous eDNA metabarcoding studies
- Shows importance of using:
  - Multiple sampling method

### Limitations

- Only one sample collected per site and method
- Hudson brush sample failed
- Sequencing depth varied between samples
- Possible:
  - PCR bias
  - Database limitations
- Some low-abundance taxa may not have been detected

### Future Directions

- Use diversity analyses:
  - Alpha diversity
  - Beta diversity
- Improve: Replication, Taxonomic databases, DNA purification methods

## References (Scan Me!)



## Acknowledgements

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